

Rapid Acceleration of Diagnostics: *RADx (Tech + ATP)*

Bruce J. Tromberg, Ph.D.

Director, National Institute of Biomedical Imaging and Bioengineering (NIBIB)



RADx Tech & ATP

NIH Office of the Director



Francis Collins



Rachael Fleurance



Larry Tabak



Tara Schwetz

**April 24, 2020: \$1.5B to NIH
\$500 Million to NIBIB**

April 29

RADx Tech – \$500M

Highly competitive, rapid three-phase challenge to identify the best of class for at-home or point-of-care tests for COVID-19

RADx Advanced Technology Platforms (RADx-ATP) – \$230M

Rapid scale-up of advanced technologies to increase rapidity and enhance and validate throughput – create ultra-high throughput machines and facilities

RADx Radical (RADx-Rad) – \$200M

Develop and advance novel, non-traditional approaches or new applications of existing approaches for testing

RADx Underserved Populations (RADx-UP) – \$500M

Interlinked community-based demonstration projects focused on implementation strategies to enable and enhance testing of COVID-19 in vulnerable populations



Jill Heemskerk



Bruce Tromberg

**National Institute of
Biomedical Imaging and
Bioengineering (NIBIB)**

Tech/ATP Team Leads: Tiffani Lash, Todd Merchak, Taylor Gilliland, Kate Egan, Mike Wolfson, Doug Sheeley, Gene Civillico



\$307 M Partnership with BARDA



<https://www.nih.gov/research-training/medical-research-initiatives/radx>;

Tromberg, Collins et al. NEJM, 2020

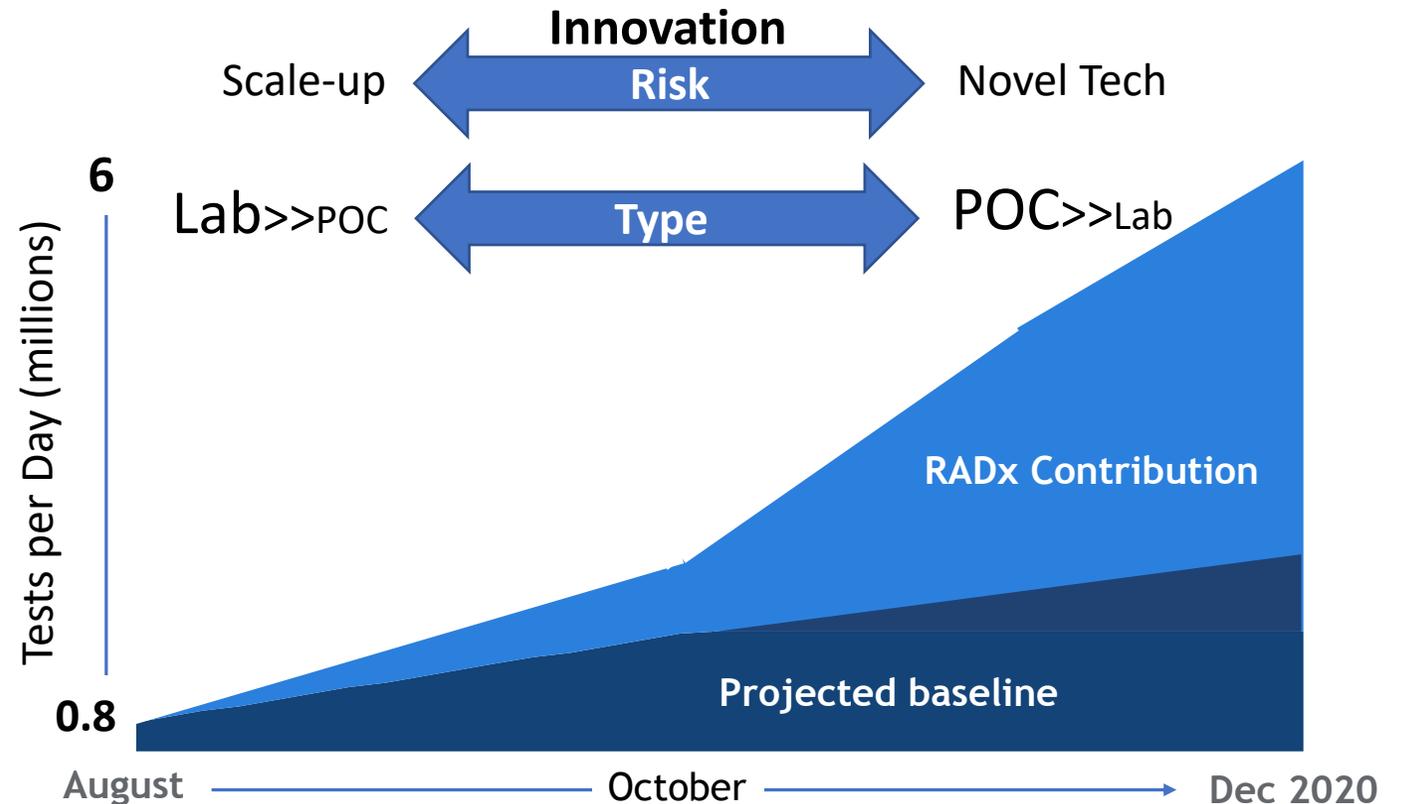
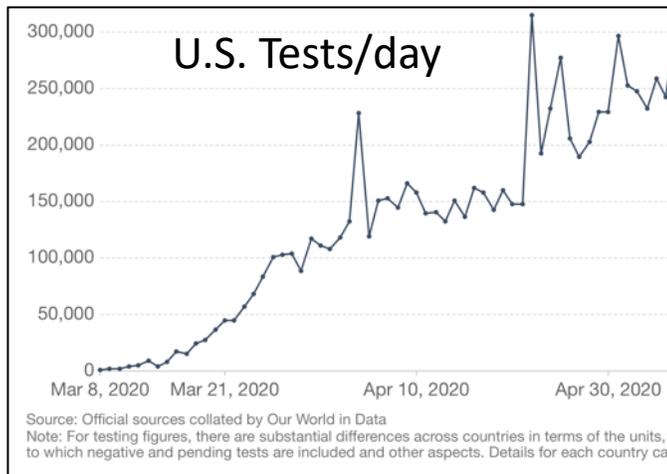
RADx Tech & ATP Goals

1) Expand COVID-19 Testing Technologies: *Number, Type and Access*

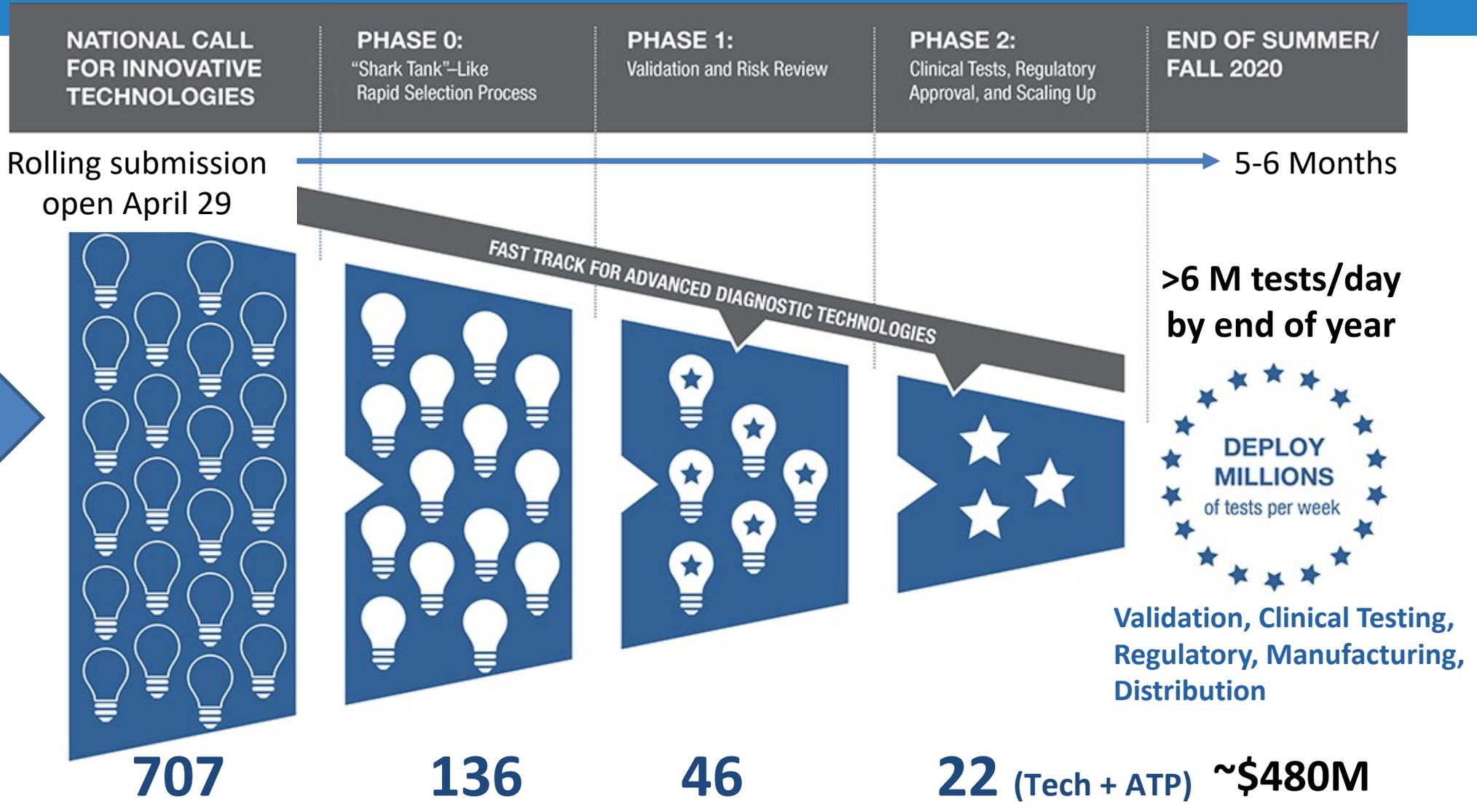
2) Optimize Performance: *Technologic and Operational; Match Community Needs*

Test Settings

- Home-based
- Point of Care (POC)
- Laboratory (CLIA, research)



RADx Tech/ATP Innovation Funnel



RADx Tech/ATP Innovation Funnel

NATIONAL CALL FOR INNOVATIVE TECHNOLOGIES

PHASE 0: "Shark Tank"-Like Rapid Selection Process

PHASE 1: Validation and Risk Review

PHASE 2: Clinical Tests, Regulatory Approval, and Scaling Up

END OF SUMMER/FALL 2020

Manufacturing Expansion Summary

5-6 Months

Applications Started

~3000

- **Type:** 17 Nucleic Acid, 5 Viral Antigen
- **Setting:** 8 POC, 3 "between", 11 Lab
- **Regulatory:** EUA → 8 lab (+1), 3 POC (+2)
- **Impact:** ~2.5M tests/day (Dec)

- **Pipeline:** 21 POC (9 NAT, 11 An, 1 VOC)

Projects in each Phase

707

136

46

22 (Tech + ATP) ~\$480M

FAST TRACK FOR ADVANCED DIAGNOSTIC TECHNOLOGIES

>6 M tests/day by end of year



validation, Clinical Testing, Regulatory, Manufacturing, Distribution

22 Manufacturing Expansion

Innovation

- 1) Separation/concentration
- 2) μ -Fluidics
- 3) Chemistries, e.g. CRISPR, NGS
- 4) Labels, Reporters
- 5) Readout Tech
- 6) Miniaturization
- 7) Automation

Tens to
100,000
tests/day

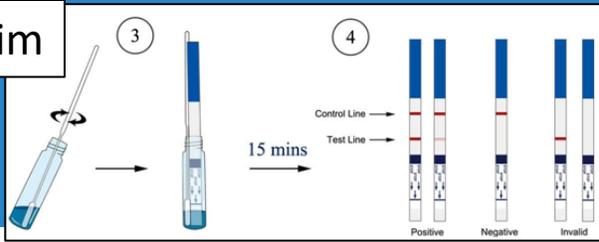
<https://www.nibib.nih.gov/covid-19/radx-tech-program/radx-tech-phase2-awards>

Visby Medical	Nucleic Acid: RTPCR	POC
MicroGEM International	Nucleic Acid: RTPCR	POC
Mesa Biotech, Inc.	Nucleic Acid: RTPCR	POC
Talis Biomedical Corp.	Nucleic Acid: Isothermal PCR	POC
MatMaCorp	Nucleic Acid: RTPCR	Lab/POC
Ubiquitome	Nucleic Acid: RTPCR	Lab/POC
Maxim Biomedical Inc	Antigen: LFA dipstick	POC
Luminostics, Inc.	Antigen: LFA	POC
Ellume USA LLC	Antigen: LFA	POC/home
Quidel Corp.	Antigen: LFA	POC
Quanterix	Antigen/microbeads	Lab
Mammoth Biosciences	Nucleic Acid: CRISPR	Lab
Flambeau Diagnostics	Nucleic Acid: Isothermal PCR	Mobile Lab
Ceres Nanosciences Inc	Nucleic Acid: Extraction	Lab
Fluidigm	Nucleic Acid: RTPCR	Lab
Broad Institute	Nucleic Acid: RTPCR	Lab
Illumina Inc	Nucleic Acid: NGS	Lab
Helix OpCo, LLC	Nucleic Acid: NGS	Lab
Ginkgo Bioworks	Nucleic Acid: NGS	Lab
Sonic Healthcare USA	Nucleic Acid: RTPCR	Lab Network
PathGroup	Nucleic Acid: RTPCR	Lab Network
Aegis Sciences	Nucleic Acid: RTPCR	Lab Network

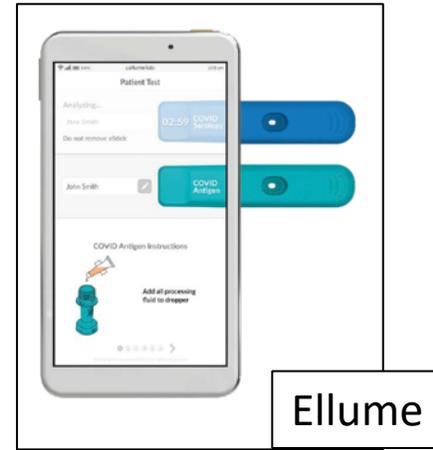


Mesa BioTech

Maxim



Quidel Sophia



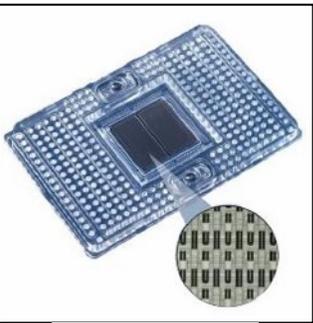
Ellume



Ubiquitome

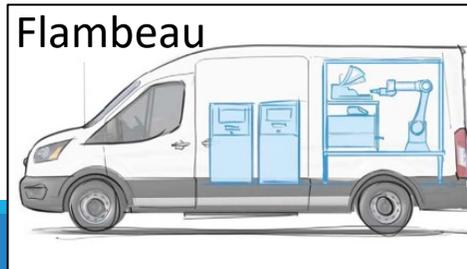


Luminostics



Fluidigm

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Sonic Healthcare USA	Nucleic Acid: RTPCR	Lab Network
PathGroup	Nucleic Acid: RTPCR	Lab Network
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Flambeau

Point-of-Care Technologies Research Network (POCTRN)

NIBIB National Network: 5-6 years for new POC technologies

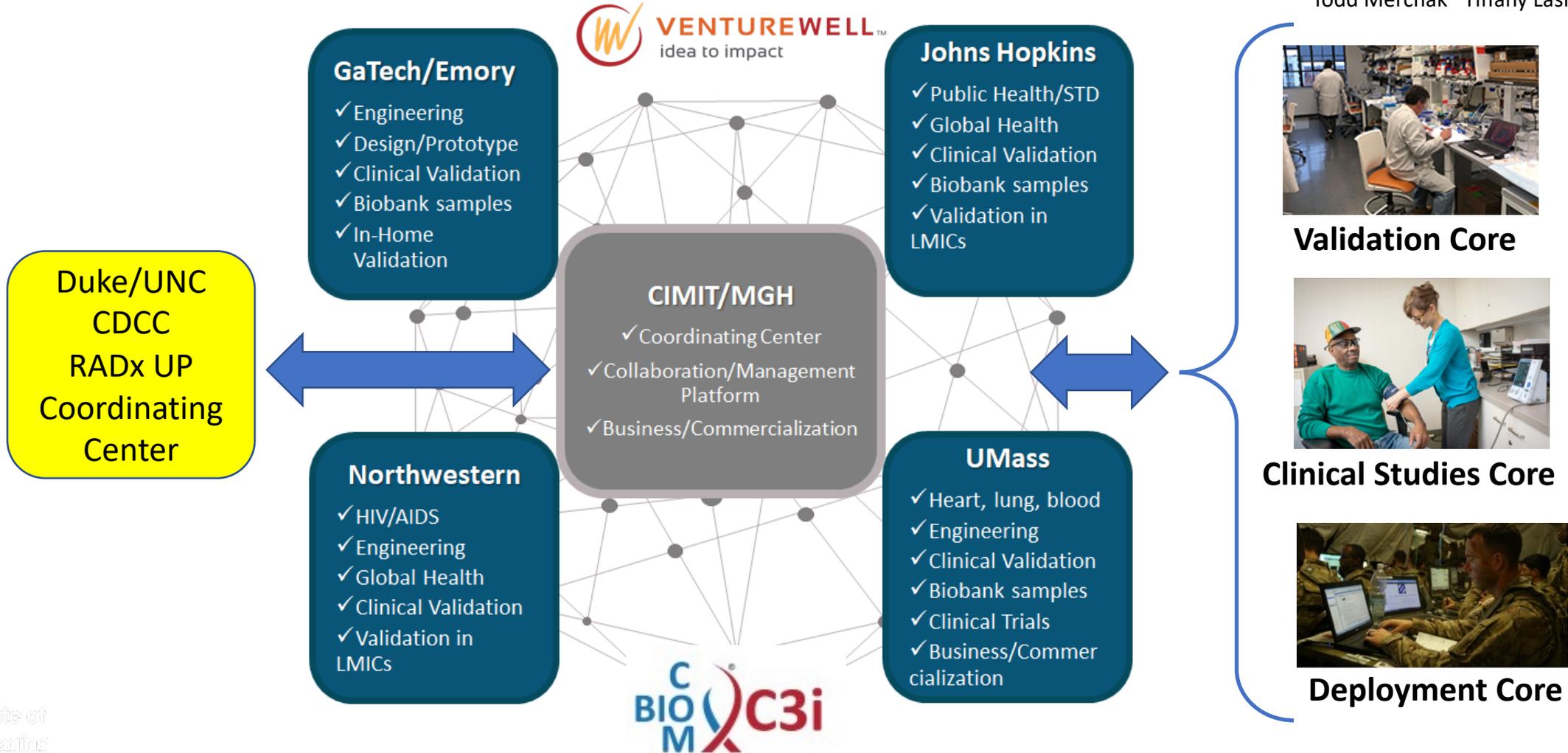
Established 2007, Expanded 2020: >1000 RADx experts & contributors

<https://www.poctrn.org>



Todd Merchak Tiffany Lash

RADx UP



RADx Leveraging NIH Proof of Concept (PoC) Network



Jodi Black



Matt McMahon

~40 early-stage RADx-tech projects



Project Funding



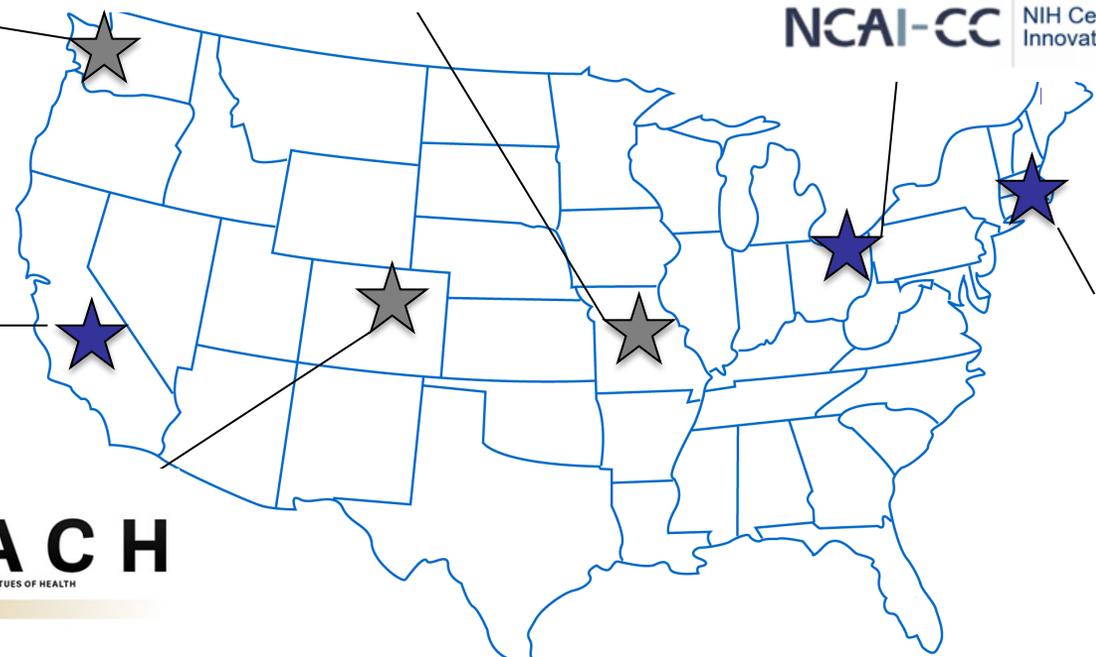
Industry Coaching and Mentoring



Training and Resources



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RADx Test Validation Core (Emory-Gtech)

15 projects complete, 11 ongoing



Wilbur Lam



Greg Martin



Oliver Brand

Feasibility

Ensure positive control (provided or commercial) is positive
Ensure negative matrix (i.e. saliva, patient sample or commercial) is negative
Ensure negative matrix spiked with live and/or inactivated SARS-CoV-2 virus is positive



Contrived samples

Verify the limit of detection (LOD) via live and/or inactivated SARS-CoV-2 virus by serial dilution using correct matrix
Test non-SARS-CoV-2 coronaviruses (test specificity/cross-reactivity)
Test different strains of SARS-CoV-2 (strain variation)



Patient samples

Test banked patient samples (adult and pediatric) with concomitant testing on reference method to determine concordance
Test prospective patient samples using collection sites **>1500 participants**
Calculate sensitivity, specificity, positive and negative predictive values with input from our biostatistical core

RADx Test Validation Core (Emory-Gtech)

15 projects complete, 11 ongoing

Feasibility

- Ensure positive control (provided or commercial) is positive
- Ensure negative matrix (i.e. saliva, patient sample or control) is negative
- Ensure negative matrix spiked with live and/or inactivated SARS-CoV-2 is positive

NIH score range: 1 (exceptional) to 9 (poor)

ACME POCT score: 2 (88% of respondents)

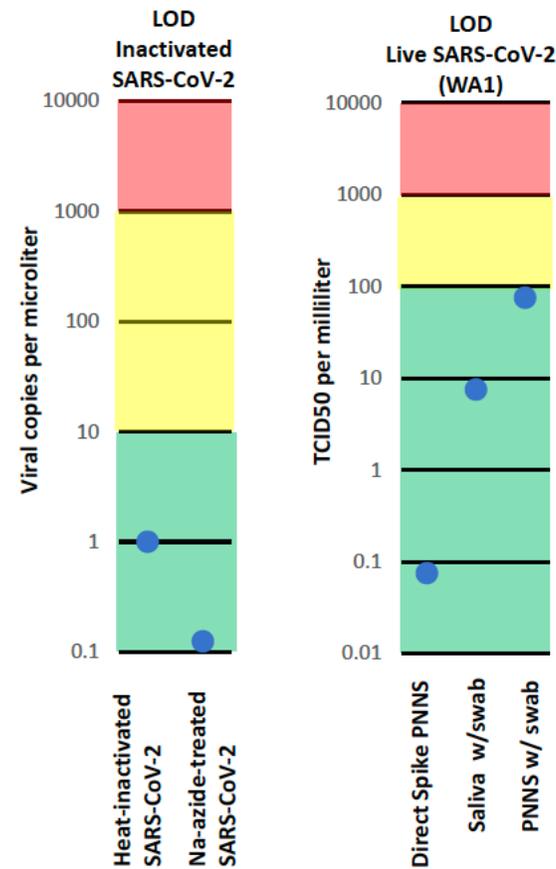
RADx Test Verification Core Recommendation: Proceed to WP2

Resume and Summary of Discussion: the RADx ACME POCT convened an internal study section on July 9th, 2020 to discuss the RADx Test Verification Core's analysis of Project #2244 in which the criteria for evaluation included: LOD, Sensitivity, Specificity, Repeatability, and Usability. The testing of this COVID-19 point-of-care (POC) PCR diagnostic test comprised of 1) LOD testing at several of our sites, including our Emory PSL 2 facility, Children's Healthcare of Atlanta clinical pathology laboratory, and laboratories in

Patient samples

- Test banked patient samples (adult and pediatric) with known status
- Test prospective patient samples using collection sites
- Calculate sensitivity, specificity, positive and negative predictive values

OVERALL SUMMARY OF RESULTS ACROSS ACME POCT SITES



Specificity	
Virus Type	Lack of Cross-Reactivity?
OC43 seasonal coronavirus	✓
MERS (heat inactivated)	✓

Prospective Clinical Results

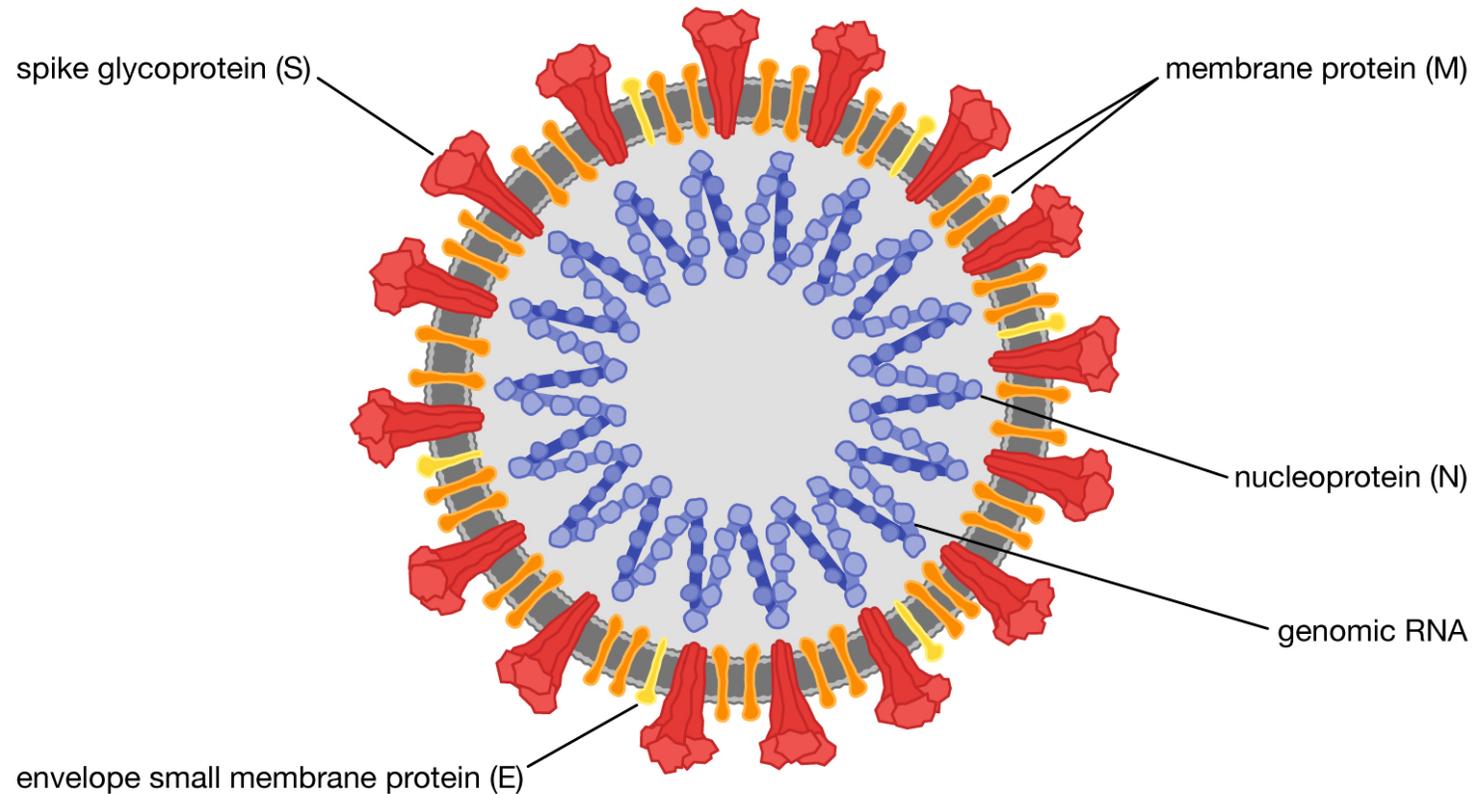
	DiaSorin RT-PCR	
	+	-
2244 +	4	0
2244 -	0	48

Sensitivity: 100%
Specificity: 100%

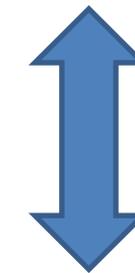
* PNNS, pooled negative nasal swab

RADx Test Validation Core (Emory-Gtech)

Challenge: Compare NAT and Antigen Test Performance



Viral Antigen Test
Lateral Flow Assay (LFA)
LOD: TCID₅₀/mL ~10³-10⁴/mL



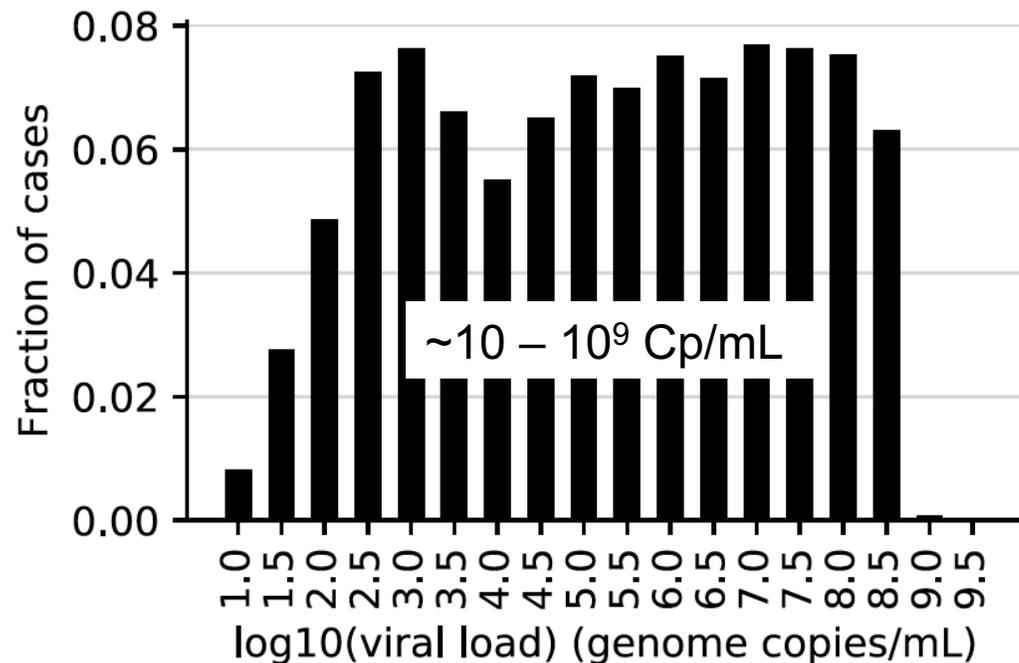
Copies/TCID₅₀ ?

Nucleic Acid Test
RTPCR (Isothermal PCR)
LOD: Cp/mL ~10²-10³/mL

Understanding Screening/Surveillance Performance

Impact of LOD and Population Viral Load on Performance

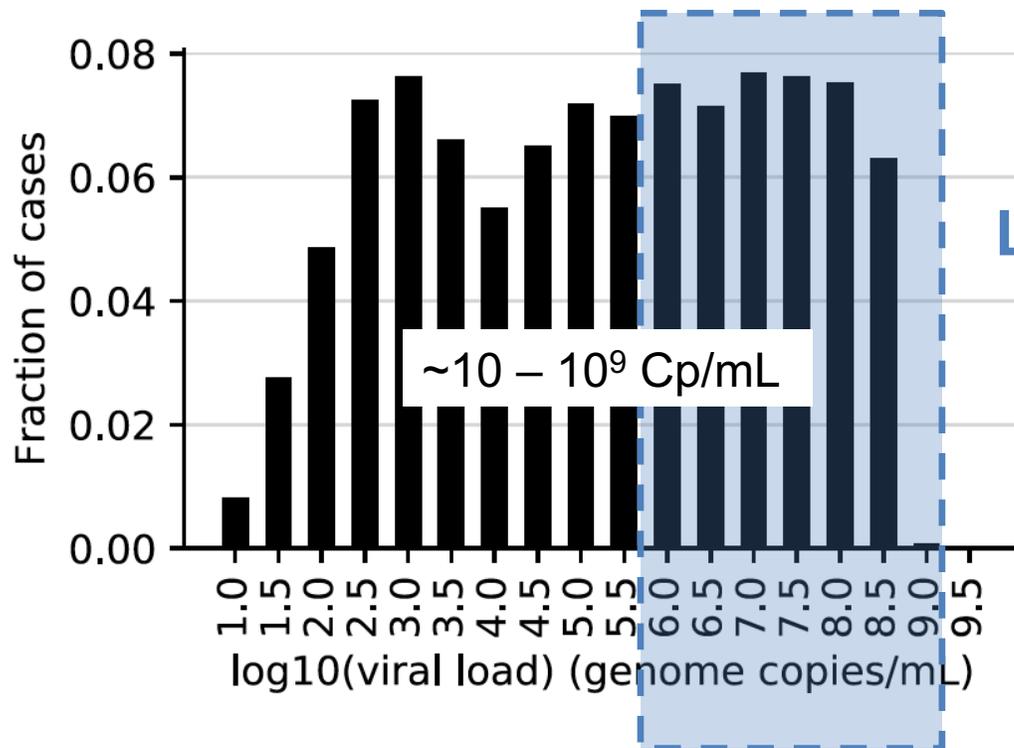
Population Viral Loads from Ct values ($n = 4774$)



Understanding Screening/Surveillance Performance

Impact of LOD and Population Viral Load on Performance

Population Viral Loads from Ct values ($n = 4774$)



LFA → Typical LOD $\sim 10^6$ Copies/mL
Sensitivity $\sim 40\%$ for all population
(symptomatic + asymptomatic)

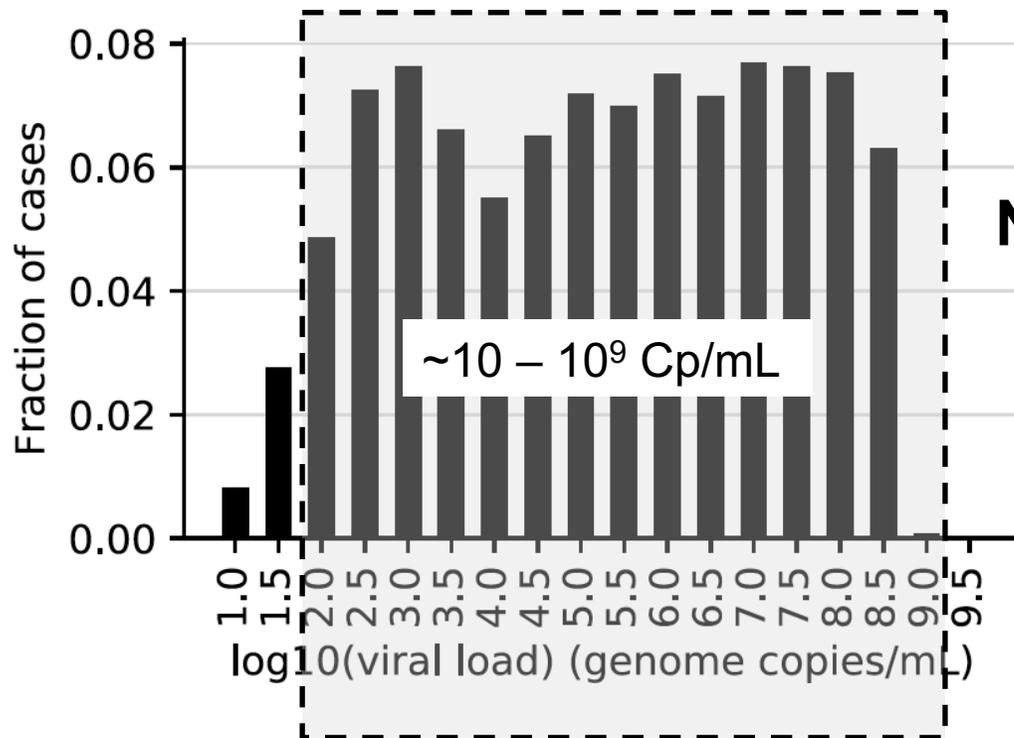
Vs.

Sens/Spec $\sim 90/95\%$ for symptomatic
population (EUA: ~ 5 days onset)

Understanding Screening/Surveillance Performance

Impact of LOD and Population Viral Load on Performance

Population Viral Loads from Ct values ($n = 4774$)

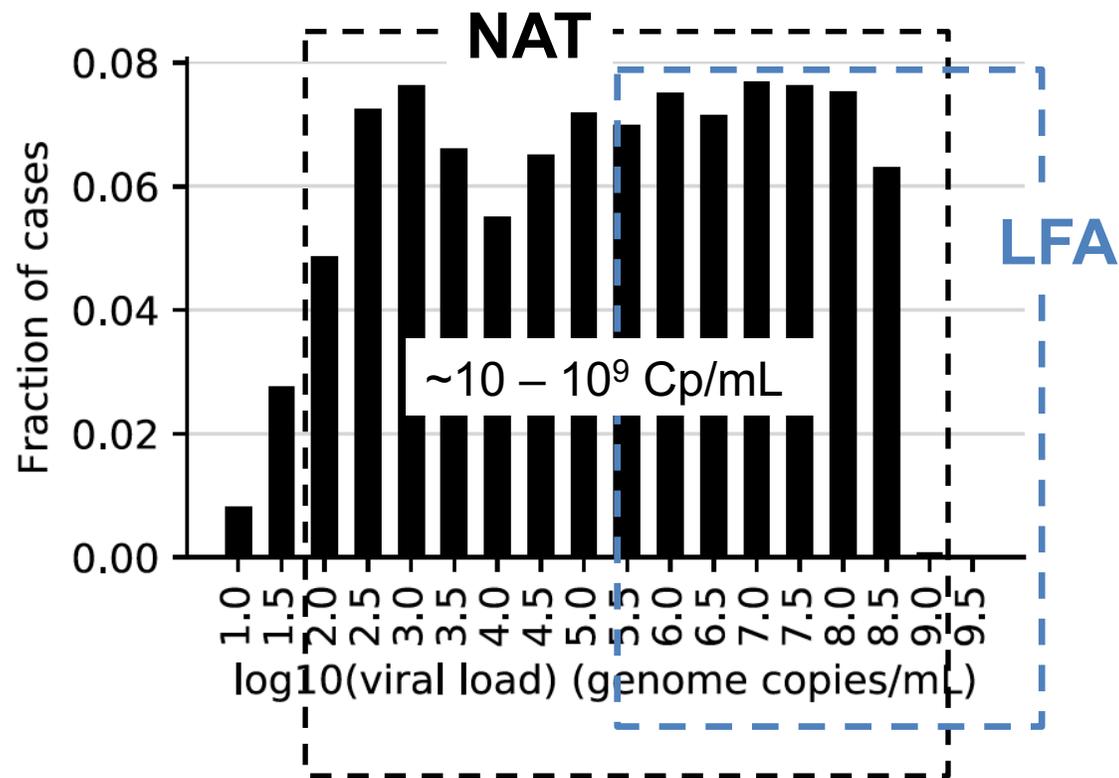


NAT → Typical LOD > $\sim 10^2$ Copies/mL
Sensitivity > $\sim 90\%$ for all population
(symptomatic + asymptomatic)

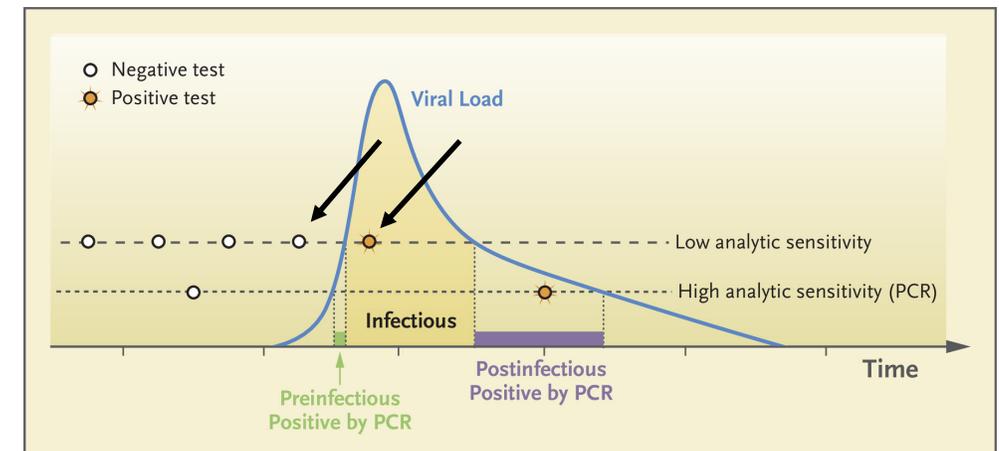
Understanding Screening/Surveillance Performance

Implications: NAT (PCR) vs LFA (An)

Population Viral Loads from Ct values ($n = 4774$)



- 1) Use LFA within $\sim 5-7$ days of symptoms
 - Elevated viral load ($>90\%$ sens, spec)
- 2) “Off Label” LFA in Asymptomatics:
 - Backup PCR w/positive in low prevalence
 - Backup PCR w/negative recently exposed
- 3) Sequential LFA tests



RADx Clinical Studies Core (UMass)

Mission: Evaluate RADx platforms that advance to Phase 2 in rigorous clinical studies w/ diverse populations and settings.

Standard Trial Design: Master protocols, powered studies (~250 subjects), device-specific amendments, accelerate regulatory review

Eureka Digital Health Platform mobile app and website, participants enter own data

Data Safety Board and Single IRB for oversight and safety monitoring

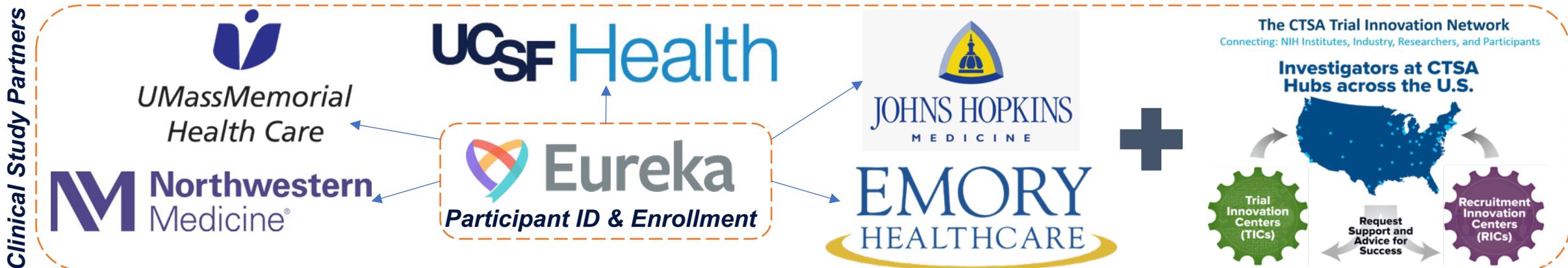
Robust Research Center Network: POCTRN core center network for enrollment (w/Practice Based Research Network and Centers for Clinical and Translational Science assisting)



Laura Gibson, MD



David McManus, MD



RADx Deployment Core (CIMIT)

Bridging NIH/USG, non-profit Foundations, Academia, and Industry

Mission

Provide support for successful commercialization and deployment of COVID-19 solutions in unique communities.

- Members: 32
- Nancy Gagliano, MD, Core Lead
- Brian Walsh, Commercialization Lead
- Sreeram Ramakrishnan, Data Solutions Lead
- Susan Moreira, Deployment Lead



Nancy Gagliano, MD

www.poctrn.org

RADx webinars, tools

Current Highlights

- Supply Chain continues to be core challenge
- Development of Testing Model has received international recognition
- User communities need end-to-end solutions to deploy COVID testing
- Design-a-thon scheduled to develop data solutions

“When-to-Test” modeling tool:
Match tests w/needs; evaluate
impact of risk reducing activities.



Anette Hosoi
MIT



Paul Tessier,
CIMIT/MGH

POC Comparison: *Performance Gap*

POC RTPCR



Visby Medical

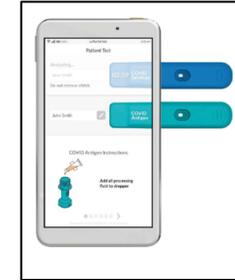


Mesa BioTech

POC An (LFA/reader)

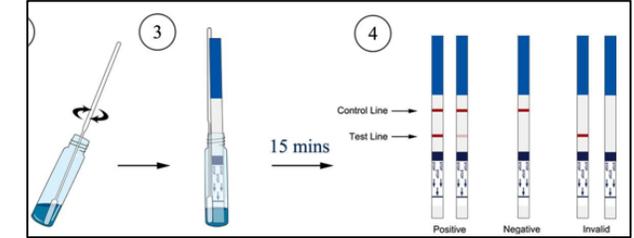


Quidel Sophia



Ellume

POC An (LFA/visual)



Maxim

Cost

\$\$\$

\$\$

\$\$

\$

Speed

~30 min

<15 min



Sens/Spec (EUA)

>90/95

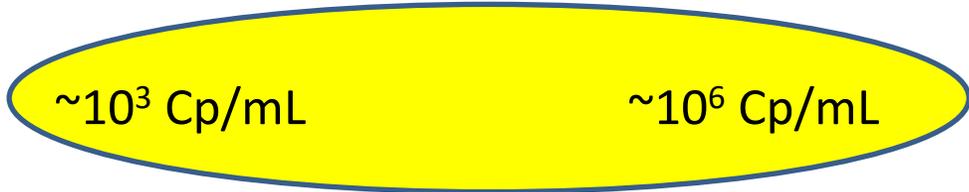
>90/95



LOD

~10³ Cp/mL

~10⁶ Cp/mL



POC Comparison: *Performance Gap*

POC RTPCR



Visby Medical

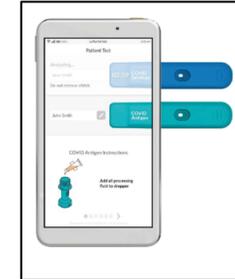


Mesa BioTech

POC An (LFA/reader)

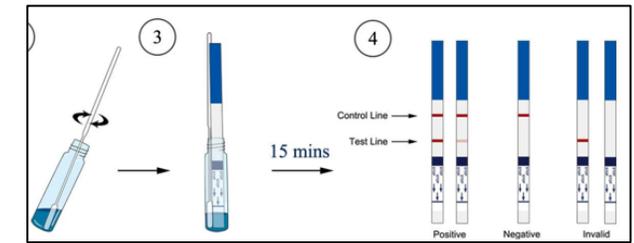


Quidel Sophia



Ellume

POC An (LFA/visual)



Maxim

Cost

\$\$

\$

\$

0.5 \$

Speed

~30 min

<15 min

Sens/Spec (EUA)

>90/95

>90/95

LOD

~10³ Cp/mL

~10⁶ Cp/mL

Tech to Bridge the Gap

- 1) Separation/concentration
- 2) μ -Fluidics
- 3) Chemistries, e.g. CRISPR
- 4) Labels, Reporters
- 5) Readout/Sensing
- 6) ? ("Background" reduction)

RADx Digital Health Networks: *Integration*

RADx POC Test

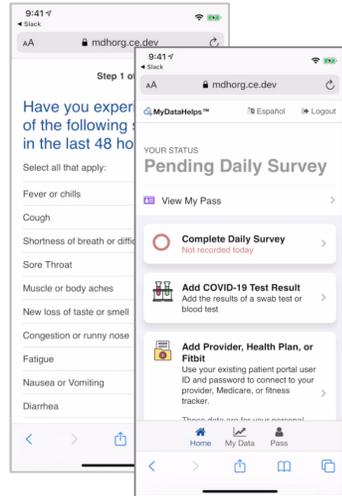


e.g. OpenRDT (Audere)

Wearables



Symptom Surveys



Cell Phone Reader



GATES foundation

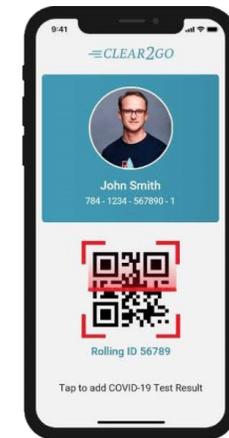
Digital Contact Tracing



EHR & Claims



Proof of Health Status



Andrew Weitz
NIBIB



Jason Levine
NCI

RADx Tech/ATP Summary

RADx Tech/ATP:

Accelerating innovation, Multiple platforms, Millions tests/day

Link NVBL to RADx network?

Implementation Challenge:

- Standard Medical Diagnostics: *accurately detect/diagnose disease in individuals*
- **COVID Paradox: rapidly assess +/- of disease in asymptomatic populations: Home?**
- Barriers: *Economic, cultural, regulatory*

Technical Challenges:

- Match tests w/biology of infection; needs of user communities (*what/when/how often?*)
- Bridge Tech performance gap: *improve performance in low prevalence settings*
- Integrate: *tests, models, interventions, Apps/data, for personal and PH management*